

A Comparative Performance Survey on Microarray Data Analysis Techniques for Colon Cancer Classification

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ABSTRACT

The genetic information of any human beings is very helpful in cancer diagnosis. DNA microarray technology has enabled us to handle thousands of genes simultaneously. cDNA and Affymetrix microarray are the microarray technologies. The microarray data analysis can be done in supervised or unsupervised learning methods. Hierarchical clustering, k-means algorithms are widely used for clustering. As Curse of dimensionality is main challenge for microarray, Feature selection techniques are used. The classification accuracy depends on the feature selection technique used. In proposed work, feature selection techniques implemented are Signal-to-Noise ratio, Information Gain and Fishers criteria. SVM and KNN classifiers are built. The comparative results of performance accuracies are generated. The SVM classifier outperforms with fishers criteria and KNN outperforms with SNR.

General Terms

Cancer Classification

Keywords

Microarray, cancer, genes, feature selection, classification

1. INTRODUCTION

Microarray is a powerful technique that allows monitoring the expression of tens of thousands of different genes in a small sample simultaneously [1]. This technique makes use of surfaces like glass slides, silicon chips on which sequences from thousands of different genes from a sample are covalently attached to a fixed location (probes) [2].

The microarray technologies currently available are cDNA microarray and affymetrix array. cDNA microarrays are small glass slides(or nylon membranes) on which double stranded DNA is spotted. Usually each spot on chip represents a gene. In cDNA microarray two samples a reference (a normal tissue) verses test sample (a malignant tissue) are used. This pair of two cDNA samples is independently copied from corresponding mRNA populations with reverse transcriptase enzyme. Distinct fluorescent molecules Cyanine5 (cy5) red and Cyanine3 (cy3) green are used for labeling. These two labeled cDNA samples are then pooled and hybridized to the array. The relative expression level of a particular gene is determined by measuring the densities at both fluorescence wavelengths and then calculating the ratios of fluorescence intensities (cy5/cy3) [2].

Affymetrix array is another technology also known as oligonucleotide array places thousands of gene-specific oligonucleotides (called probes) synthesized directly on silicon chip using a photolithographic technology. This synthesis uses in situ (on chip) light directed chemistry to

build up many thousands of oligonucleotide probes (each probe 25 bp long). Each gene has 15-20 pair of probes synthesized on the chip. Each pair of probes has two oligonucleotides perfect match (PM) and mismatch (MM). PM (reference sequence) is perfectly complementary to a specific region of gene or EST(Expression sequence tag) and MM (one base change) is identical to perfect match probes except for a single middle base pair mismatch [3][4].

The affymetrix scanner generates a TIFF (Tagged Image File Format) image of scanned array and store in DAT file. Image analysis is performed on DAT file and CEL file is generated which stores the probe level expression data (probe intensities). CDF (Chip Description File) describes which probe goes in which probe set. The computation of gene expression values for requires both CEL and CDF file. This computational process requires three steps background correction, normalization and summarization. Microarray Analysis Suite5.0 (MAS 5), Robust Multichip analysis (RMA), Model Based Expression Index (MBEI), GCRMA are the algorithms used for analysis of microarray. From studies it is concluded that RMA is superior to MAS in terms of sensitivity and specificity (i.e. true and false detection rate) [5].

Microarray data analysis is facing two major challenges small number of microarray data samples are available from small number of patients and curse of dimensionality, thousands or tens of thousands of genes. Many genes from the dataset contain irrelevant information for the accurate classification of disease. There is high redundant and irrelevant information in dataset needed to be removed. The extraction/selection of most relevant gene set is very important for accurate classification results. Feature selection techniques reduce large dimensional data set into smaller gene set capable of distinguishing between infected and normal samples [1][6].

The microarray classification process is a two stage process: feature selection and classification. There are three widely known feature selection approaches: filter, wrapper and embedded methods. Filter methods rank genes according to certain intrinsic characteristics of gene expressions with the class label. Filters are of two types, univariate or multivariate. The univariate filter considers intrinsic properties of each feature individually ignoring feature dependencies. T-statistics, Chi-square, Signal-to-Noise ratio (SNR), Information Gain (IG), Gain Ratio (GR) are univariate filters. The multivariate filters take feature dependencies into consideration. Correlation based feature selection (CFS) is a multivariate filter. Wrapper methods interact with the classifier while gene ranking. In embedded methods the gene selection process is embedded in constructing classifier [7]. Some of the classification techniques are Support Vector

Machine (SVM) [1] [6], K- Nearest Neighbors (KNN) [1], Naïve Bayes, Neural Network, and Decision Tree (DT).

In this work, for first stage of classification authors have used three feature selection techniques, Signal-to-Noise Ratio (SNR), Information Gain (IG) and Fisher Criteria for extracting important predictive genes. In second stage of classification 2 powerful classifiers Support Vector Machine (SVM) and K Nearest Neighbor (KNN) are built. 4 publically available gene expression datasets are used.

This manuscript is organized as follows: Section 2 summarizes the related work. The proposed system is described in section 3. In Section 4 the experimental results and analysis are reported. Section 5 is summary and conclusion.

2. RELATED WORK

The process of classification and clustering are generally considered to be similar, the only difference is classification is a supervised learning process (i.e. it is known that which training tuple belongs to which class) and clustering is unsupervised learning process (i.e. class label of training tuple is not known).

2.1 Clustering

Cluster analysis on gene expression values allows grouping of all genes or samples identical to each other based on some

criteria. Cluster is collection of objects similar to each other and dissimilar objects belongs to another cluster. These clustered known genes can be used for prediction of function of unknown genes. Hierarchical clustering, K-means, Self-organizing Map (SOM) are the clustering algorithms [4].

2.1.1 Hierarchical Clustering

There are two approaches of hierarchical clustering: Top-down approach (divisive clustering) and Bottom-up approach (agglomerative clustering). Gene expression data is clustered using agglomerative approach. In this approach initially each gene expression profile is assigned to a single cluster. The distance between every couple of cluster is measured using any distance measure i.e. Euclidean, Manhattan or correlation distance measure. The two closest clusters are merged, each time the distance matrix is updated to take this cluster merging into account. The widely used methods for calculation of inter cluster distance are: single linkage (minimum distance), complete linkage (maximum distance), average linkage (average distance), centroid linkage (distance between cluster centroids). This is an iterative process until one cluster is left. This gives a tree structure where height of the branches is proportionate to distance between the clusters [4].

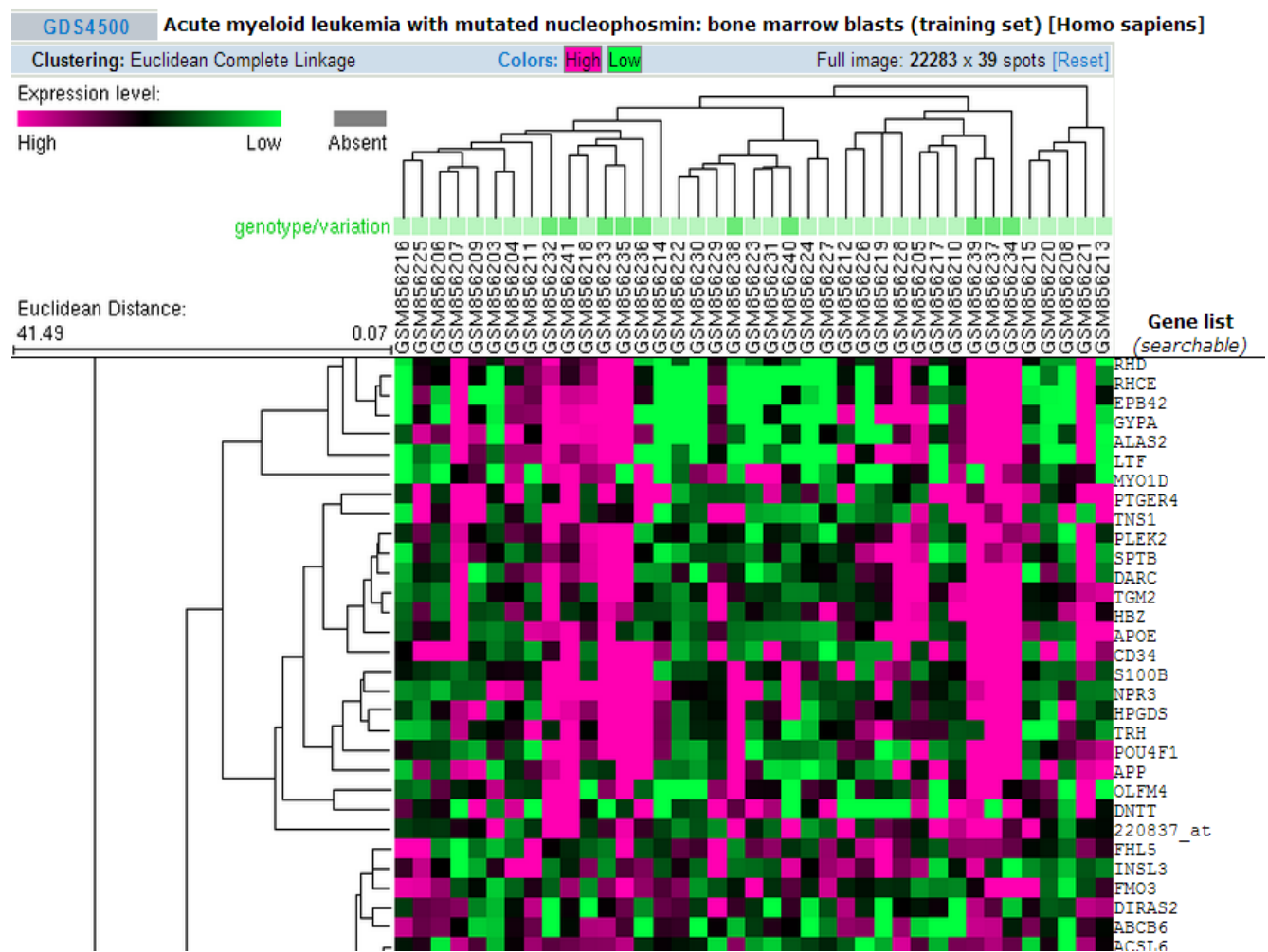


Fig 1: Hierarchical Clustering on AML GDS4500

Figure1. shows the hierarchical clustering applied on the Acute Myeloid Leukemia (AML) dataset of 39 samples having 22283 gene expression values available on NCBI GEO public data repository GDS 4500. NCBI GEO allows clustering on microarray dataset using K-means, Hierarchical clustering algorithm using average, complete and single linkage method and Euclidean, Pearson Correlation and Uncentered Correlation distance measures. K-means allows clustering for values of k ranging between 2 to 15. The column indicates clustering among samples and row indicates clustering among gene expression values [8]. There are many application software available which allows various operations to be performed on microarray dataset. Bioconductor is one of the most famous open source application software available.

2.1.2 K-Means Clustering

In K-Means clustering each gene expression value belongs to exactly one predefined clusters K. This is an iterative process; center of cluster is computed iteratively followed by assignment of gene expression value to the cluster with the closest cluster center.

2.1.3 Self-organizing Map

In SOMs there is a predefined geometry of nodes, two dimensional grid one node for each cluster. The gene expression value is mapped to the node which is closest to it.

2.2 Feature Selection

Feature selections techniques are used to identify smaller subset of most relevant features which can be used for accurate classification. The feature relevance score of each feature is calculated, and features having low score are removed. The top ranked genes are used to build the classifier. The feature selection techniques are given below.

2.2.1 Signal-to-Noise (SNR) Ratio

In the SNR method, consider a dataset S consisting of m expression vectors, $X^i = (x_1^i, \dots, x_n^i)$, $1 \leq i \leq m$ where m is the number of patient samples and n is the number of genes measured. For each gene g_i , we calculate the mean μ_i and standard deviation σ_i . It is the ratio of difference of mean of the two classes to the summation of standard deviations of two classes.

$$SNR = \frac{\mu_i - \mu_j}{\sigma_i + \sigma_j}$$

μ_i and μ_j denote the average expression value of i^{th} gene over all samples in normal and tumor case respectively. σ_i and σ_j denote standard deviation of i^{th} gene over all samples in normal and tumor case respectively.

2.2.2 Information Gain (IG)

Information Gain is computed using entropy value. Entropy of Y is

$$H(Y) = - \sum_{y \in Y} p(y) \log_2(p(y))$$

$p(y)$ is the marginal probability density function for random variable Y. Conditional entropy of Y after observing X is

$$H(Y|X) = - \sum_{x \in X} p(x) \sum_{y \in Y} p(y|x) \log_2(p(y|x))$$

$p(y|x)$ is conditional probability of y given x [9][10]. The information gained about Y after observing X is

$$IG = H(Y) - H(Y|X)$$

Information gain is symmetrical measure,

$$IG = H(Y) - H(Y|X) = H(X) - H(X|Y)$$

2.2.3 Fisher's Criteria

In Fisher's Criteria the gene ranking is done using following equation

$$fisher(g) = \frac{(m_1(g) - m_2(g))^2}{s_1^2(g) + s_2^2(g)}$$

m_1 and m_2 denote the mean expression value of g^{th} gene across all samples in tumor and normal case respectively. s_1 and s_2 denote standard deviation of g^{th} gene across all samples in tumor and normal case respectively [11].

2.3 Classification

2.3.1 Support Vector Machine (SVM)

Support Vector Machine (SVM) is a kind of supervised learning methods for classification. SVM is a popular and powerful classification technique. The main objective of SVM is to construct optimal hyperplane where two classes are linearly separable or a set of hyperplanes in a high dimensional feature space. An optimal hyperplane is one having maximum margin of separation between different classes. The data points closest to the hyperplane are considered as support vectors. Consider γ is the width of the margin, then all the data points on or within this margin will form the subset of support vectors. The data points located at distance more than $\gamma/2$ from the separating hyperplane are ignored. The support vectors play a greater role in classifying the test samples. The complexity and accuracy of SVM classifier is based on number of support vectors rather than the dimensionality of the dataset. The two parameters in hyperplane equation are w and b. The separating hyperplane equation is written as

$$w \cdot X + b = 0$$

where w is a weight vector and b is a scalar often referred as a bias.

Given training set of instance-label pairs (x_i, y_i) , $i=1, \dots, n$, where x_i are training sample with y_i class label and n is the number of samples in training set. The linear SVM classifier requires solving the following optimization problem

$$\min_{w,b,\varepsilon} \frac{1}{2} ||w||^2 + C \sum_{i=1}^n \varepsilon_i$$

subject to

$$y_i(w \cdot X_i + b) \geq 1 - \varepsilon_i, \varepsilon_i \geq 0, \forall i = 1, \dots, n$$

where C is the SVM sensitivity parameter, and ε is a slack variable. The dual formation of above optimization problem is

$$\min_{\alpha} = \frac{1}{2} \sum_{i=1}^n \sum_{l=1}^n \alpha_i \alpha_l y_i y_l X_i X_l - \sum_{i=1}^n \alpha_i$$

subject to

$$\sum_{i=1}^n y_i \alpha_i = 0, 0 \leq \alpha_i \leq C, \forall i = 1, \dots, n \text{ and } \forall l = 1, \dots, n$$

where α_i is Lagrange multiplier, obtained by training SVM [1][6][12].

2.3.2 K-Nearest Neighbor

KNN is a lazy learning algorithm where k nearest neighbors' are used for classification. There is no explicit training phase on the training dataset. For classification of the test sample all the training data is used. At the time of classification, it computes the distance between test data and training data elements using distance measures. Euclidean distance, Manhattan distance, Cosine distance, or Correlation distance are the distance matrix. It finds the k closest training points of test sample to classify test sample. The class labels of these k closest points are used to find class of test sample.

The proper functioning of KNN depends on selection of parameter k i.e. number of nearest neighbors chosen to assign class label to test data and distance matrix used. Euclidean distance measure is most widely used in KNN [1] [13] [14].

Consider two vectors x_i and x_j such that $x_i = (x_{i1}, x_{i2}, \dots, x_{in})$ and $x_j = (x_{j1}, x_{j2}, \dots, x_{jn})$. The Euclidean distance between x_i and x_j is given by

$$d(x_i, x_j) = \sqrt{\sum_{r=1}^n (x_{ir} - x_{jr})^2}$$

and the Manhattan distance is given by

$$d(x_i, x_j) = \sqrt{\sum_{r=1}^n |x_{ir} - x_{jr}|}$$

3. PROPOSED METHOD

In this paper the authors have comparatively studying and analyzed three feature selection and two classifications

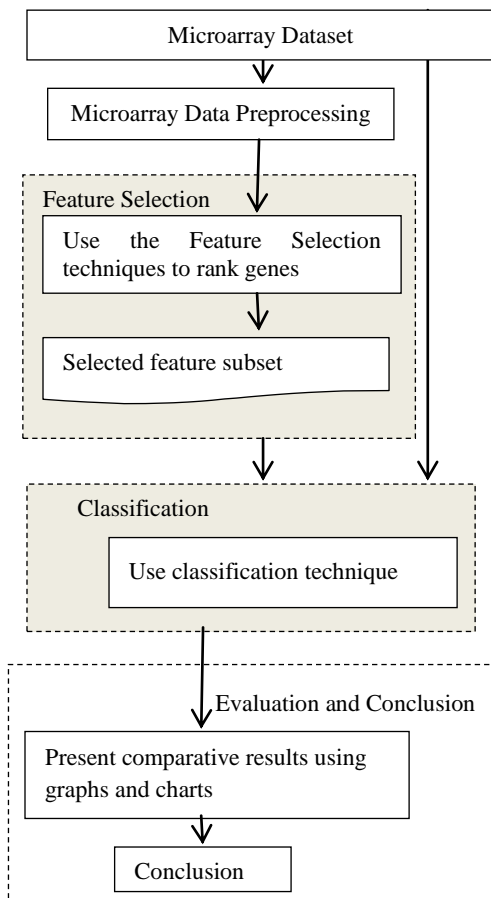


Fig 2: Proposed microarray data analysis system

techniques for microarray data analysis for accurate classification of cancer.

The workflow of the proposed microarray data analysis system for cancer classification is shown in Figure 1. The feature selection techniques used are: Signal to Noise Ratio (SNR), Information Gain (IG) and Fishers Criteria. Support Vector Machine (SVM) and K-Nearest Neighbor (KNN) classifiers are built. The features extracted are used to build classifier.

Many online repositories are there which make the biomedical data set available which include microarray gene expression value, protein informative data, and genomic sequence data. The proposed method is using the dataset made available on Kent Ridge Bio-medical Data Set Repository.

The Colon cancer dataset is used. The dataset contains expression of 2000 genes across 62 samples collected from Colon Tumor patients. Among these samples, 40 are tumor biopsies (labeled as “positive”) from tumors and 22 are normal biopsies (labeled as “negative”) from healthy parts of the colons of the same patient.

There are four possible outcomes for a given classifier and an instance. (1) True positive (TP) if the positive instance is classified as positive, (2) False negative (FN) if positive instance is classified as negative (3) True negative (TN) if the negative instance classified as negative (4) False positive (FP) if negative instance is classified as positive [10]. The accuracy of a classifier is the fraction of the correctly classified samples to all samples.

$$Accuracy = \frac{TN + TP}{TN + FN + FP + TP}$$

4. RESULTS AND DISCUSSION

In this work, a two class dataset (normal, tumor), the colon cancer dataset is used. The dataset gives different percentage of accuracies for different feature selection and classification techniques. Table 1 to Table 6. shows classification accuracies of SVM and KNN classifier with Signal- to Noise ratio (SNR), Information Gain(IG) and Fisher Criteria (FC) feature selection techniques when top 5,25,50,100,150,200 genes are selected.

Table 1. Accuracies (%) with 5 top ranked genes selected

Classifier	SNR	IG	Fisher Criteria
SVM	85.48	98.38	98.38
KNN	88.7	64.51	93.54

Table 2. Accuracies (%) with 25 top ranked genes selected

Classifier	SNR	IG	Fisher Criteria
SVM	83.87	79.03	95.16
KNN	99.9	83.87	99.9

Table 3. Accuracies (%) with 50 top ranked genes selected

Classifier	SNR	IG	Fisher Criteria
SVM	79.03	87.09	87.07
KNN	99.9	87.09	87.07

Table 4. Accuracies(%) with 100 top ranked genes selected

Classifier	SNR	IG	Fisher Criteria
SVM	80.64	88.8	93.54
KNN	99.9	93.54	99.9

Table 5. Accuracies(%) with 150 top ranked genes selected

Classifier	SNR	IG	Fisher Criteria
SVM	75.8	96.71	91.93
KNN	99.9	99.9	99.9

Table 6. Accuracies(%) with 200 top ranked genes selected

Classifier	SNR	IG	Fisher Criteria
SVM	75.8	53.22	91.93
KNN	99.9	93.54	99.9

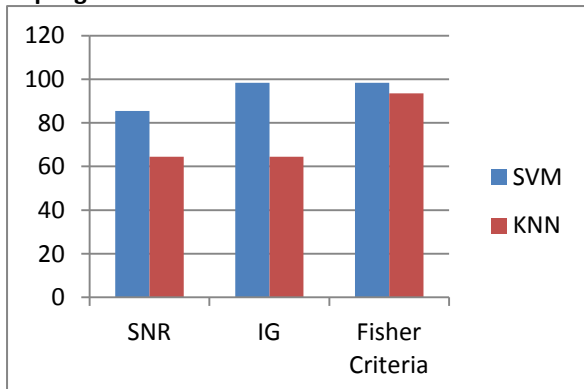
The performance of SVM classifier is good with Fishers Criteria. Fisher's Criteria gives most relevant feature subset for classification than SNR and IG. The good performance of SVM does not depend on the dimensionality of the dataset but on the support vectors selected for classification. Because of support vector the SVM turns out to be the best performance classifier.

The performance of KNN classifier is good with Signal-to-Noise ratio (SNR) than IG and FC.

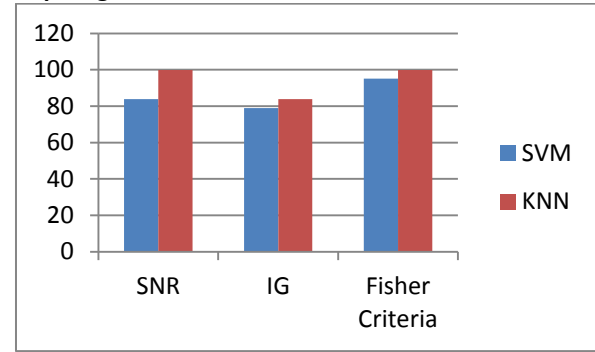
Figure 2. shows the performance results of the classification experiment using three feature selection techniques, SNR, IG and FC and considering top 5,25,50,100,150,200 ranked genes (refer Table 1 to Table 6.)

Figure 3. shows the performance results of the classification experiment considering top 5,25,50,100,150,200 ranked genes for classification. Results show that SVM gives 98.38 % of accuracy when implemented using Fishers criteria with top 5 ranked genes. KNN classifier gives 99.9 % of accuracy when implemented using Signal-to-Noise ratio (SNR) with top 25 features selected.

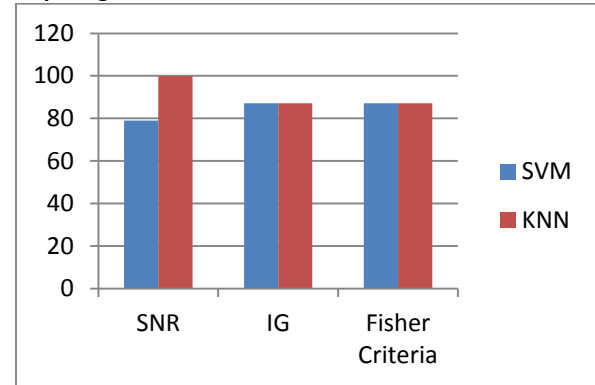
Top 5 genes selected



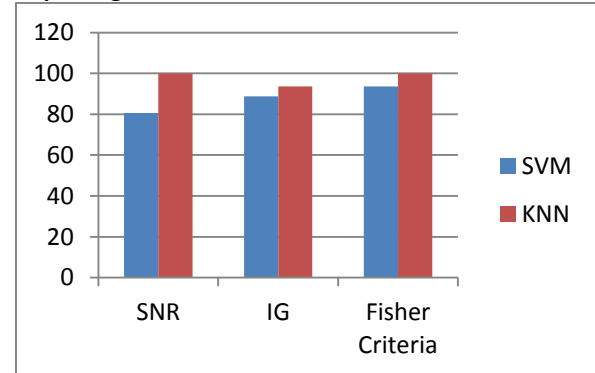
Top 25 genes selected



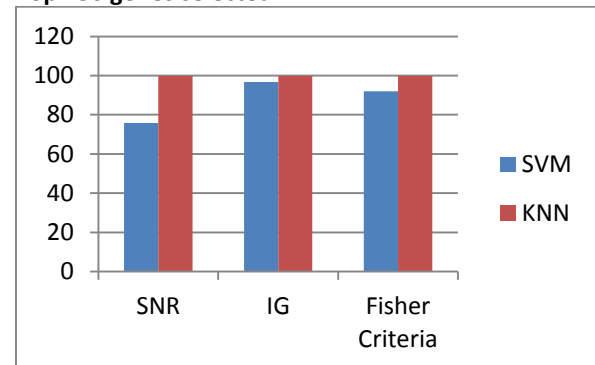
Top 50 genes selected



Top 100 genes selected



Top 150 genes selected



Top 200 genes selected

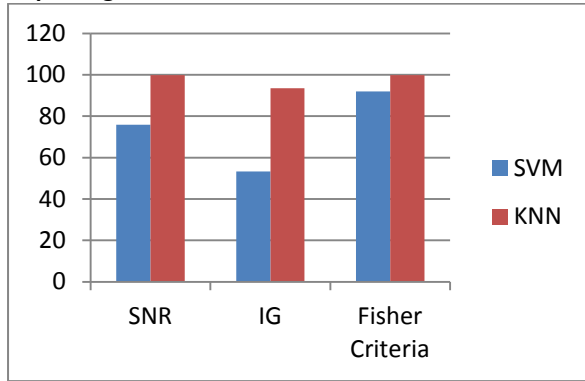
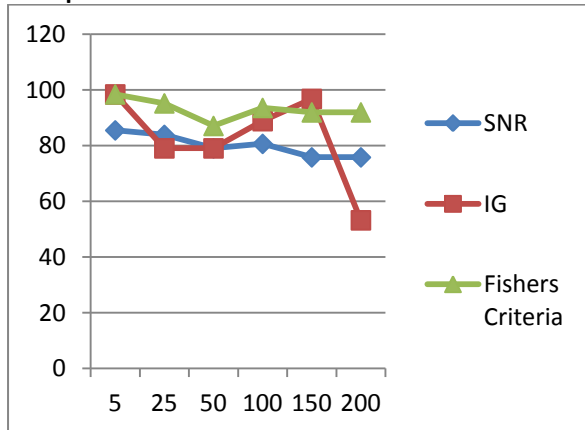


Fig 2: Performance result of classification with gene selection using SNR, IG and FC with different number of top ranked feature selection (genes)

SVM performance



KNN performance

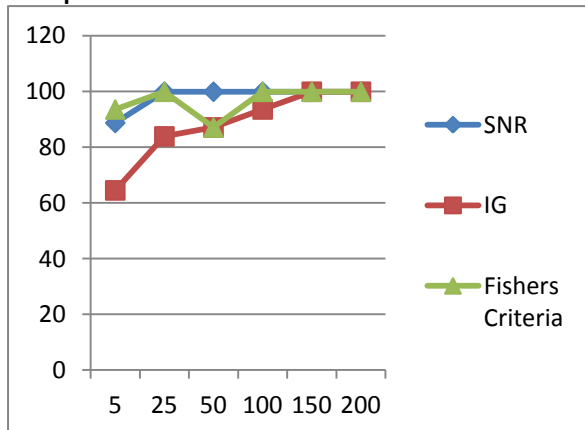


Fig 3: Performance result of classification with gene selection using SNR, IG and FC with top 100 genes

5. CONCLUSION

Microarray data can be analyzed by unsupervised learning (clustering) or supervised learning (classification). Hierarchical clustering is most widely used clustering algorithm for gene expression values.

The accuracy of classifier depends on the feature selection method used for finding the most relevant and informative features. The classifier is built using these extracted features. A good feature selection technique is one which extracts the

most relevant features for classification. The accuracy of classifier varies depending upon the number of top ranked features selected for classification. SVM best performs with Fishers Criteria using 5 features selected and KNN gives better performance with SNR feature selection technique using 25 feature selected.

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8. AUTHOR’S BIOGRAPHY

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