

In Silico Studies of Fullerene C₆₀ with Zebrafish Proteins Prostacyclin Synthase and S100z Calcium Binding Protein

Nateshan Anil
Assistant professor,
Sreenidhi institute of
science and
technology,
Yamnapet,
Ghatkesar,
Hyderabad

S.S.Vutukuru
Professor&Head,
Sreenidhi
institute of science
and, Technology
Ghatkesar.
Hyderabad

K.S.R.Sivasai
Professor,
Sreenidhi institute
of science and
technology,
Yamnapet,
Ghatkesar
Hyderabad

Jayasree
Ganugapati
Associate Professor
Sreenidhi institute of
science and
technology,yamnap
t,
Ghatkesar, Hyderabad

ABSTRACT

Nanotechnology offers many societal benefits and this have prompted the rapid growth of engineered nanomaterials. Fullerene (C₆₀) due to its unique properties has become an important nanomaterial in biomedical and biotechnological applications. Once in water, Fullerene forms stable suspended aggregates and thus become bioavailable to aquatic biota. The fate and transport of fullerene in the aquatic environment is poorly understood. Little data are available on the molecular interactions of fullerene with native proteins of zebra fish (*Danio rerio*) which is a universally accepted experimental model. In this study, we made an attempt to assess the binding mode of fullerene with two key zebrafish proteins viz. Prostacyclin synthase (cytochrome P450 8A1) and S-100Z Calcium binding protein using Autodock 4.0. The data indicates that Fullerene potentially binds to the active sites of both the proteins and this may induce conformational changes in the native protein structure there by altering the function, which might have a toxic effect to fish in survival. Further in vivo studies are required to evaluate the toxic impact on the expression and function of above proteins.

Keywords:

Fullerene (C₆₀), Prostacyclin synthase, S-100Z Calcium binding protein, Docking, Autodock 4.0.

1. Introduction

Fullerene (C₆₀) is made up of hollow carbon molecules arranged in a cage of interlocking pentagons and hexagons like the patches on the soccer ball with flexible chemical reactivity (1,2). Fullerene (C₆₀) with 7.2 Å in diameter is similar in size to steroid hormones or peptide alpha helices, and thus considered as ideal molecules to serve as Ligand for enzymes and receptors (3). Fullerene is shown to exhibit bioactivity in antibacterial, neuroprotection and DNA cleavage etc (4-8). Although data is still emerging on the toxicity of engineered nanomaterials, research done so far indicates that

Fullerene is capable of inhibiting the growth of prokaryotic cells, certain enzymes like glutathione reductase and neuronal nitric oxide synthase (9-11). The unique properties of fullerenes have also risen to another concern, their potential risks to biological systems. Research on fullerene is critical

given the high production of these materials and their possible harmful ecological effects.

Little data are available on the molecular interactions of fullerene with native proteins of Zebrafish (*Danio rerio*) which is universally accepted experimental model. Zebrafish (*Danio rerio*) a small tropical fish which lives in rivers of India, Northern Pakistan and Nepal and Bhutan in South Asia is widely used model in many areas of biological research. Furthermore, *Danio rerio* is an excellent system for chemical toxicity testing (12-13).

Prostacyclin synthase (CYP8A1) (5.3.99.4) is located on chromosome 6 which catalyses the rearrangement of Prostaglandin H₂ (PGH₂) to prostaglandin I₂, this endoplasmic reticulum membrane protein catalyzes the conversion of prostaglandin H₂ to Prostacyclin (Prostaglandin I₂), a potent vasodilator and inhibitor of platelet aggregation. An imbalance of Prostacyclin contributes to the development of myocardial infarction, stroke, and atherosclerosis in humans (Fig. 1) (14). S-100 protein is a family of low molecular weight protein found in vertebrates characterized by two calcium binding sites of the helix-loop-helix ("EF-hand type") S100 is normally present in cells derived from the neural crest (Schwann cells, melanocytes, and glial cells), chondrocytes, adipocytes, myoepithelial cells, macrophages, Langerhans cells, dendritic cells and have been implicated in a variety of intracellular and extracellular functions. S100 proteins are involved in regulation of protein phosphorylation, transcription factors, Ca²⁺ homeostasis, the dynamics of cytoskeleton constituents, enzyme activities, cell growth and differentiation, and the inflammatory response (15, 16).

This study is an attempt to evaluate the binding mode of Fullerene C₆₀ with two key zebrafish proteins namely Prostacyclin synthase Cytochrome (P₄₅₀8A1 5.3.99.4) and S100Z calcium binding protein using AutoDock 4.0.

2. MATERIALS AND METHODS

Docking Studies

To study the nature of interactions and binding mode of zebra fish proteins and fullerene. Docking was performed using Auto dock 4.0

2.1 Preparation of protein

PDB (17) is a single worldwide archive of structural data of biological macromolecules, established in Brookhaven national laboratories (BNL) in 1971. It contains structural information of the macromolecules determined by x-ray crystallography's and NMR methods. The Zebra fish protein structures of Prostacyclin synthase (Cytochrome P450 8A1) with PDB ID 3B98 (Figure2) and, S100Z calcium binding protein with PDB ID 2Y5I (Fig. 2) were retrieved from PDB.

2.2 Preparation of Ligand

The 3D structure of Fullerene C60 (Ligand) (Fig. 3) with ID110185 was retrieved from ChemSpider (18). ChemSpider is a free chemical structure database providing fast access to over 25 million structures, properties and associated information. By integrating and linking compounds from more than 400 data sources, ChemSpider enables researchers to discover the most comprehensive view of freely available chemical data from a single online search. It is owned by the Royal Society of Chemistry. ChemSpider builds on the

collected sources by adding additional properties, related information and links back to original data sources. ChemSpider offers text and structure searching to find compounds of interest and provides unique services to improve this data by curation and annotation and to integrate it with users' applications.

3. Active Site Analysis:

Q-site Finder (19), an online tool which uses hydrophobic probes, was used to predict possible binding sites.

4. Auto dock 4.0

Autodock (20) is the first docking package to model the ligand with full conformational stability. The package consists of two sequentially applied programs, Autogrid and Autodock. Autogrid is initially used to calculate noncovalent energy of interaction between the rigid part of the receptor and a probe atom that is located at various points of the lattice. Autodock's main strengths are receptor flexibility, blind docking, Precalculated grid maps on a binding site, free energy scoring function based on linear regression analysis, the AMBER force field and a large set of Protein ligand complexes with known inhibition.

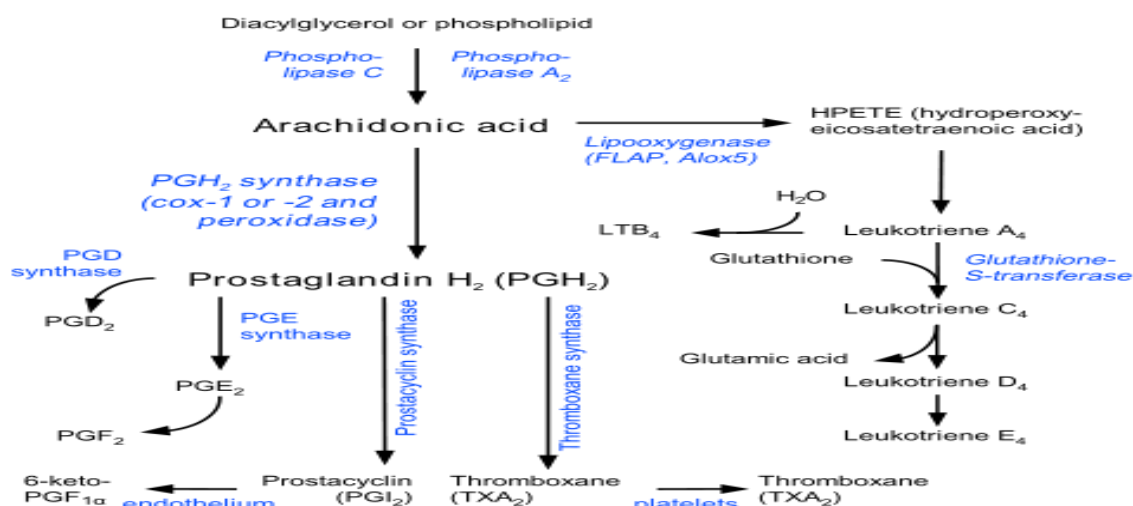


Fig. 1: Cyclooxygenase, lipoxygenase and epoxygenase pathways leading to the formation of eicosanoids from arachidonic acid. Fullerene binds to prostacyclin synthase and may inhibit formation of prostacyclin (PGI₂)



Fig. 2: A. Prostacyclin Synthase (PDB ID 3B98); B. S-100 Z Calcium Binding Protein (PDB ID 2Y5I)

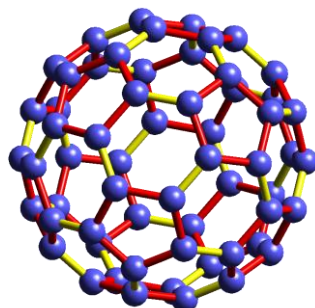


Fig.3: Fullerene (C₆₀)

3.0 RESULTS AND DISCUSSION

Prostacyclin synthase and S100z calcium binding protein are important enzymes related to metabolism in Zebrafish. Prostacyclin synthase is an endoplasmic reticulum membrane protein that catalyzes the conversion of prostaglandin H₂ to prostacyclin (Prostaglandin I₂), a potent vasodilator through the arachidonic acid pathway. Our study indicates that Fullerene (C₆₀) binds to the active site of Prostacyclin synthase using Autodock 4.0. The binding energy was found to be -11.96 Kcal/mol as shown in Table I. The interactive residues of prostacyclin synthase with fullerene are depicted in Table 2.

Fullerene interacts with the residues, PHE126, VAL273, THR274, ASN277, CYS421, GLY423, which are similar residues of Prostacyclin synthase, binding to selective substrates which is depicted in Table 2, where the residues are bolded. The interactive residues of prostacyclin synthase with fullerene are depicted in structure view; this is represented in Fig 4A. Hence, we speculate that Fullerene could inhibit the activity of Prostacyclin synthase, leading to inhibition of vasodilation and ultimately affecting the survival of fish in the long term.

S100 protein is a low molecular weight protein found in vertebrates, characterized by two calcium binding sites of helix-loop-helix ("EF-hand type") involved in regulation of protein phosphorylation, transcription factors, Ca²⁺ homeostasis, the dynamics of cytoskeleton constituents. Our study also indicates that Fullerene (C₆₀) binds to the active site of S100Z Calcium binding protein using Autodock 4.0. The binding energy was found to be -3.78 Kcal/mol as shown in Table I. The interactive residues of S100Z Calcium binding

protein with Fullerene are depicted in Table 2. Fullerene interacts with the residues ARG 97, GLN 93, LEU 82, etc. The interactive residues of S100Z Calcium binding protein with fullerene are depicted in structure view; this is represented in Fig 4B. Hence, we speculate that Fullerene could inhibit the activity of S100Z Calcium binding protein, which might have a toxic effect on fish.

Our docking study also suggests that Fullerene binds to Prostacyclin synthase with more specificity compared to S100Z, based on the binding energies (Table 1), and the number of identical residues (Prostacyclin synthase has seven residues common between the substrate and Fullerene compared to S100Z, which has three residues) binding between the selective substrate and Fullerene.

(Table 2) in the active site. Because of this, the production of prostacyclin synthesis may decrease and lead to a loss of vasoprotective activity due to an increase in vasoconstriction, aggregation, and platelet activation in fish.

S100Z Calcium Binding, the binding of Fullerene may have an effect on the regulation of protein phosphorylation, transcription factors, Ca⁺⁺ homeostasis, which may have a deleterious effect on the activity of fish in nature and environment.

Table. 1: Energy value (Kcal/mol) of Proteins obtained by Auto dock 4.0 upon binding with Fullerene

SNo	Protein	Binding energy Kcal/mol
1	Prostacyclin synthase	-11.96
2	S100z calcium binding protein	-3.78

Table. 2: Residues found in the binding pocket of respective proteins using Q-site finder and residues in the active site region

S.No	Enzyme	Residues found in the binding pocket of proteins using Q-site finder	Fullerene bind to residues in the active site region
1	Prostacyclin synthase (3B98)	ALA122, PHE126 ,MET138,LEU171,PHE172,THR174,GLY175,VAL179,TRP245,TRP273, VAL273,THR274 ,GLN275,GLY276, ASN277 ,ALA278,GLY279,ALA281,LEU,321,LEU324,TRP414, CYS421,PRO422,GLY423 ,PHE426,ALA427,ALA431,ILE465	PHE126,VAL273,THR274,ASN277, CYS421, GLY423,
2	S100Z Calcium binding protein (2Y5I)	PHE90, GLN93 ,GLN94, ARG97 ,SER98LEU38,LEU42,PHE45,LEU46,LEU54,ILE58,LEU78, LEU82	LEU82, GLN 93, ARG97.

* **Bolded Residues** are identical residues with interacting with selective substrate and Fullerene for prostacyclin synthase and S100Z calcium binding protein in the active site.

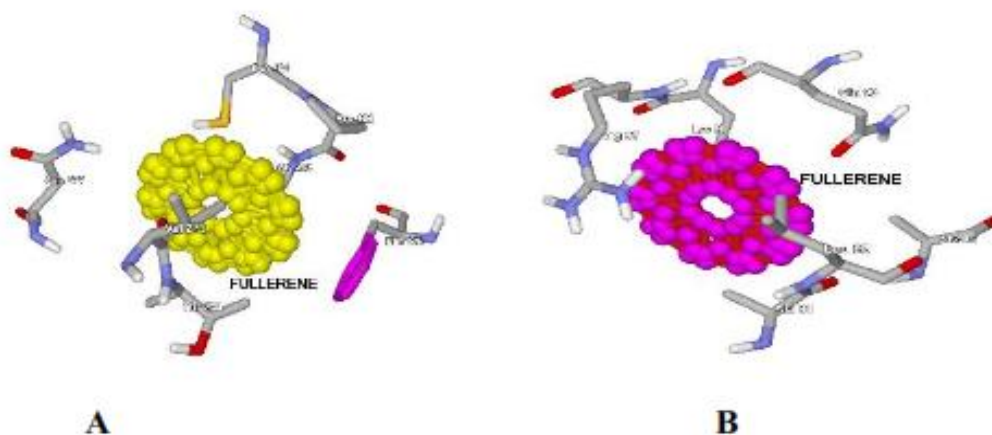


Fig .4: A. Fullerene C₆₀ interaction with prostacyclin synthase 3B98, The ligand which is in ball and stick model shown in yellow color and the active site residues of protein are labelled and shown in stick model;

B. Fullerene interaction with S 100 Z calcium binding Protein, The ligand which is in ball and stick model shown in pink color and the active site residues of protein are labeled and shown in stick model

5.0 CONCLUSION

Fullerene (C₆₀) binds to same residues as selective substrates for Prostacyclin synthase and S100Z Calcium binding protein in the active sites suggests that Fullerene could inhibit the activity of Prostacyclin synthase and S100Z activity. For further risk assessment of fullerene toxicity repeated studies following various animal models, methods of administration, and mechanisms of toxicity should be conducted.

6.0 ACKNOWLEDGMENTS

My sincere thanks to Dr. Rajashekar and Rajitha who helped me in doing work and also Sreenidhi institute of science and technology for providing Facilities to do my work .

7.0 REFERENCES

- [1] Kratshmer, W., Lamb, L.D., Fostiropoulos,K., andHuffman, D. Solid C60 :a new form of carbon. Nature (1990) 347, 354-358 (1990)
- [2] Kroto,H.W.,Health,J.R.,Brein,S.C.,Curl,R.F.,andSmalley ,Buckminstrfullerene.Nature(1985) 318,162.
- [3] Nanomedicine: Fullerene and Carbon Nanotube Biology. Stephen R. Wilson. C Sixty, Inc. 11 Chatham Street. Chatham, NJ 07928.
- [4] Bosi,S.,Da Ros,T.,Spalluto,G., and Prato,M. Fullerene Derivatives: An Attractive tool for biological applications. E.J.Med.Chem. (2003) 38,913-923.
- [5] Dugan,L.L., Turetsky, D.M., Du, C., Lobner,D.,Wheeler,M.,Almli,C.R.,Shen,C.K.,Luh,T.Y.C hoi,D.W.,andLin,T.S.Carboxyfullereneas,neuroprotective agents.Proc.Natl.Acad.Sci.U.S.A.949434-9439.(1997)
- [6] Jin,H.,Chen,W.Q.,Tang,X.W.,Chiang,L.Y.,Yang,C.Y.,Schloss,J.V,Polyhydroxylated,fullerenol as Glutamate receptor antagonists and neuroprotective agents.J.Neurosci. Res.62,600-607(60)
- [7] Tokuyama,H.,Yamago,S.,Nakamura,E.,Shiraki,T.,and Sugiura, Photoinduced biochemical activityofFullerenecarboxylicacid.J.Am.Chem.Soc.115,7 918-7919(1993)

- [8] Kim, J. E., and Lee, M., (2003) Fullerene inhibits beta-amyloid peptide aggregation. *Biochem. Biophysics. Res. Commun* 303, 576-579.
- [9] Mashino, T., Okuda, K., Hirota, T., Hirobe, M., Nagano, T. & Mochizuki, M. (1999). Inhibition of E. coli growth by fullerene derivatives and inhibition mechanism. *Bioorganic and Medicinal Chemistry Letters* 9, 2959–62
- [10] Mashino, T., Okudo, Hirota, T., HIROBE, M., Nagano, T., and Mochizuki, M. (2001) Inhibitory effect of fullerene derivatives On glutathione reductase . *Fullerene science . Technol .* 9 , 191- 196 .
- [11] Wolff, D. J., Barberi, C. M., Richardson, C.F., Schuster, D.I., and Wilson, S.r. (2002) Trisamine C (60)-fullerene inhibit neuronal nitric oxide Synthase by acting as highly potent calcium modulator antagonists. *Biochem., Biophys. Res. Commun* 299, 130-141.
- [12] Kerrie L Taylor, Nicola J Grant, Nicholas D Temperley and E Elizabeth Patton *Cell Communication and Signaling* 2010, Small molecule screening in zebrafish: an *in vivo* approach to identifying new chemical tools and drug leads.
- [13] Adrian J. Hill, Hiroki Teraoka, Warren Heideman,, and Richard E. Peterson1, Zebrafish as a Model Vertebrate for Investigating Chemical Toxicity, *TOXICOLOGICAL SCIENCES* 86(1), 6–19 (2005).
- [14] Yi-Ching Li , Chia-Wang Chiang, Hui-Chun Yeh , Pei-Yung Hsu, Frank G. Whitby , Lee-Ho Wang and Nei-Li Chan , Structures of Prostacyclin Synthase and Its Complexes with Substrate Analog and Inhibitor Reveal a Ligand-specific Heme Conformation Change .The journal of biological chemistry vol 283 no 5 pp 2917-2926 february 1, 2008
- [15] Olga V. Moroz ,Igor B. Bronstein, Keith S. Wilson . The Crystal Structure of Zebrafish S100Z: Implications for Calcium-Promoted S100 Protein Oligomerisation. *Journal of molecular biology*, vol 411 issue 5, 2 September 2011 1072- 1082.
- [16] Donato R, Functional roles of S 100 proteins ,calcium binding proteins of the EF- hand type . *Biochem biophysics acta* 1999 Jul 8, 1450: 191-231.
- [17] H.M. Berman, K. Henrick, H. Nakamura (2003): Announcing the worldwide Protein Data Bank. *Nature Structural Biology* 10 (12), p. 980 15
- [18] ChemSpider www.chemspider.com.
- [19] Laurie AT, Jackson RM (2005). Q-SiteFinder: an energy-based method for the prediction of protein-ligand binding Sites. *Bioinformatics*, 21: 1908-1916 Pubmed.
- [20] Morris, G. M., Goodsell, D. S., Halliday, R.S., Huey, R., Hart, W. E., Belew, R. K. and Olson, A. J. (1998), Automated Docking Using a Lamarckian Genetic Algorithm and Empirical Binding Free Energy Function *J. Computational Chemistry*, 19: 1639-1662.
- [21] Kratshmer, W., Lamb, L.D., Fostiropoulos, K., and Huffman, D. Solid C60 :a new form of carbon. *Nature* (1990) 347, 354-358 (1990)
- [22] Kroto, H.W., Heath, J.R., Brein, S.C., Curl, R.F., and Smalley, B. Buckminsterfullerene. *Nature* (1985) 318, 162.
- [23] Nanomedicine: Fullerene and Carbon Nanotube Biology. Stephen R. Wilson. C Sixty, Inc. 11 Chatham Street. Chatham, NJ 07928.
- [24] Bosi, S., Da Ros, T., Spalluto, G., and Prato, M. Fullerene Derivatives: An Attractive tool for biological applications. *E.J. Med. Chem.* (2003) 38, 913-923.
- [25] Dugan, L.L., Turetsky, D.M., Du, C., Lobner, D., Wheeler, M., Almlı, C.R., Shen, C.K., Luh, T.Y., Choi, D.W., and Lin, T.S. Carboxyfullerenes, neuroprotective agents. *Proc. Natl. Acad. Sci. U.S.A.* 94, 9434-9439. (1997)
- [26] Jin, H., Chen, W.Q., Tang, X.W., Chiang, L.Y., Yang, C.Y., Schloss, J.V. Polyhydroxylated fullereneol as Glutamate receptor antagonists and neuroprotective agents. *J. Neurosci. Res.* 62, 600-607 (2000)
- [27] Tokuyama, H., Yamago, S., Nakamura, E., Shiraki, T., and Sugiura, Photoinduced biochemical activity of Fullerene carboxylic acid. *J. Am. Chem. Soc.* 115, 7918-7919 (1993)
- [28] Kim, J. E., and Lee, M., (2003) Fullerene inhibits beta-amyloid peptide aggregation. *Biochem. Biophysics. Res. Commun* 303, 576-579.
- [29] Mashino, T., Okuda, K., Hirota, T., Hirobe, M., Nagano, T. & Mochizuki, M. (1999). Inhibition of E. coli growth by fullerene derivatives and inhibition mechanism. *Bioorganic and Medicinal Chemistry Letters* 9, 2959–62
- [30] Mashino, T., Okudo, Hirota, T., HIROBE, M., Nagano, T., and Mochizuki, M. (2001) Inhibitory effect of fullerene derivatives On glutathione reductase . *Fullerene science . Technol .* 9 , 191- 196 .
- [31] Wolff, D. J., Barberi, C. M., Richardson, C.F., Schuster, D.I., and Wilson, S.r. (2002) Trisamine C (60)-fullerene inhibit neuronal nitric oxide Synthase by acting as highly potent calcium modulator antagonists. *Biochem., Biophys. Res. Commun* 299, 130-141.
- [32] Kerrie L Taylor, Nicola J Grant, Nicholas D Temperley and E Elizabeth Patton *Cell Communication and Signaling* 2010, Small molecule screening in zebrafish: an *in vivo* approach to identifying new chemical tools and drug leads.
- [33] Adrian J. Hill, Hiroki Teraoka, Warren Heideman,, and Richard E. Peterson1, Zebrafish as a Model Vertebrate for Investigating Chemical Toxicity, *TOXICOLOGICAL SCIENCES* 86(1), 6–19 (2005).
- [34] Yi-Ching Li , Chia-Wang Chiang, Hui-Chun Yeh , Pei-Yung Hsu, Frank G. Whitby , Lee-Ho Wang and Nei-Li Chan , Structures of Prostacyclin Synthase and Its Complexes with Substrate Analog and Inhibitor Reveal a Ligand-specific Heme Conformation Change .The journal of biological chemistry vol 283 no 5 pp 2917-2926 february 1, 2008
- [35] Olga V. Moroz ,Igor B. Bronstein, Keith S. Wilson . The Crystal Structure of Zebrafish S100Z: Implications for

- Calcium-Promoted S100 Protein Oligomerisation. Journal of molecular biology, vol411 issue 5, 2 September 2011 1072- 1082.
- [36] Donato R, Functional roles of S 100 proteins ,calcium binding proteins of the EF- hand type . Biochem biophysics acta 1999 Jul 8,1450: 191-231.
- [37] H.M. Berman, K. Henrick, H. Nakamura (2003): Announcing the worldwide Protein Data Bank. Nature Structural Biology 10 (12), p. 980 15
- [38] ChemSpider www.chemspider.com.
- [39] Laurie AT, Jackson RM (2005). Q-SiteFinder: an energy-based method for the prediction of protein-ligand binding Sites. Bioinformatics, 21: 1908-1916 Pubmed.
- [40] Morris, G. M., Goodsell, D. S., Halliday, R.S., Huey, R., Hart, W. E., Belew, R. K. and Olson, A. J. (1998), Automated Docking Using a Lamarckian Genetic Algorithm and and Empirical Binding Free Energy Function J. Computational Chemistry, 19: 1639-1662.